

High-Speed CO₂ Ramping Technique

Rapid A/C_i Curves in Minutes

The CIRAS-4 can change CO₂ gas concentration while simultaneously and continuously recording data. Linear ramps of CO₂ concentration are easier to program and record than in previous instruments. Now that there is no longer a need to define a response script for a ramp test in the CIRAS-4, the process is faster and more streamlined than ever.

This application note describes the simple steps needed to set up the CIRAS-4 to run a linear ramp experiment and record data. Data are plotted to illustrate the linear ramp capability, followed by a description of the post processing of the data to generate A vs. C_i curves from the ramped gas exchange data.

Set Up the CIRAS-4 and Record Data

The following steps illustrate the streamlined process to create A-C_i curves with the CIRAS-4 in *minutes*.

1. **Perform a *Stored Diff Bal Calibration*** to allow the CIRAS-4 to have accurate offset information applied continuously throughout the linear ramp: Install fresh desiccants and a new CO₂ cartridge in the CIRAS-4 and allow it to warm up for 30 minutes. Navigate to **Setup > Calibration > Stored Diff Bal** and tap **Start**. The CIRAS-4 will step through six CO₂ levels between 0 and 2000 μmol mol⁻¹, and six H₂O levels between 0 and 100% of ambient humidity, performing a Diff Balance at each level. It then computes a regression analysis of the resulting offsets that will be applied for every CO₂ concentration in real time. The Stored Diff Balance Calibration takes 12 minutes and must be allowed to continue to completion. Tap the **Accept** button to complete the process.
2. **Set Zero Diff Bal Mode:** Navigate to **Control > General**. Tap the Zero Diff Balance Mode button, select **Zero/Stored Diff Bal** and **Save**.
3. **Set the Ramp Criteria:** Ramps in the CIRAS-4 always begin at the current control setting of CO₂ as shown on the Measurement CO₂ tile and return to that original CO₂ value when the ramp is complete. A typical ramp for a C₃ leaf might be an increasing ramp, starting at 100 μmol mol⁻¹ and ending at 1100 μmol mol⁻¹ with a ramp rate of 180 μmol mol⁻¹ per minute for a total experiment time of 5.3 minutes. To set these ramp criteria, first navigate to **Measure > Photosynthesis** and tap the CO₂r tile. Set the starting CO₂ for the ramp to 100 μmol mol⁻¹. Next, navigate to **Control > Ramp**. Set the Direction to **UP**, the Step to 180 μmol mol⁻¹ per minute, the Upper Limit to 1100 μmol mol⁻¹.and PARI to 1200 μmol m⁻¹ s⁻¹.
4. **Set the Recording Options for Ramp Processing:** Navigate to the main measurement screen **Measure > Photosynthesis**. Tap **Record Options**. Tap the **Data File Name** white box and enter a file name. Tap the **Enable Ramp** toggle so that a **1** is visible, indicating that it is enabled. The Interval will default to one second. Tap **Return**.
5. **Perform the Empty Leaf Ramp:** The first ramp recording creates the baseline trace to characterize the time response of the system and stores the data for subsequent post processing (*See Tip #2 on page 5*). With a closed cuvette, tap **Start** on the command menu (located to the right) to begin the ramp experiment. Use **Measure > Graph** or **Measure > Photosynthesis** to switch between graph and tile views while the ramp is ongoing. Once the linear ramp is complete, an additional 30 seconds of data are recorded and the CO₂ control will return to the initial Lower Limit.
6. **Perform the Ramp Experiment with a Leaf:** Tap **Record Options** on the command menu and enter a new file name as in step 4. Tap **Return**. Now with a leaf in the PLC4 Universal Leaf Cuvette chamber, tap **Start**. If one watches the plot of A vs. time for the ramp with the leaf, it becomes clear when CO₂ saturation occurs, at which time the ramp can be terminated by selecting **Stop**. You can also let it run until completion and an additional 30 seconds of data will be recorded. Either way, once the ramp is completed, the CO₂ control returns to the initial setting and is then ready for the next ramp experiment.

7. *Transfer data files to a PC to begin post processing:* Tap **Transfer Files** on the command menu. Insert a USB flash drive into either slot on the back of the CIRAS-4 console. Highlight the data files recorded in the previous steps and tap the → button to move the files to the USB drive.

8. *Post Processing:* The CIRAS-4's Stored Diff Balance capability makes post processing very simple—all done in Excel simply by adding 3 new columns to the standard CIRAS-4 output file.

To begin, open the file for the empty cuvette ramp in Excel. Copy the column of assimilation rates (A), which is column AB. Open the ramp file with the leaf in the cuvette, and paste the empty cuvette A rates into an empty column. Subtract the empty cuvette A rates from the rates obtained with the leaf in the cuvette. The differences are the actual assimilation rates, beginning after a brief lag period (typically at about line 20).

9. *Compute C_i:* The sub-stomatal CO₂ concentration, C_i, is now recomputed within the spreadsheet using the new actual Assimilation and the other values that have not changed (CO₂ analysis, and g_s and E):

$$C_i (\mu\text{mol mol}^{-1}) = \frac{\left[\left(g_c - \frac{E}{2} \right) \times C_{out} \right] - A}{\left(g_c + \frac{E}{2} \right)}$$

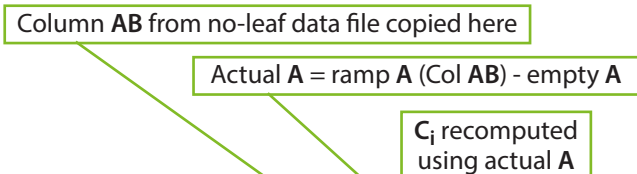
Where g_c is the total conductance to CO₂ transfer:

$$g_c (\text{mmol m}^{-2} \text{ s}^{-1}) = \left[\frac{1}{(1.585 \times r_a) + (1.37 \times r_b)} \right] \times 10^3$$

[1.585 is the diffusion ratio of CO₂ and water in air, and 1.37 is the diffusion ratio of CO₂ and water in the boundary layer.]

An Excel spreadsheet is available from PP Systems to use as a template for this calculation.

10. Plot A vs. C_i, starting at about line 20, after the linear ramp stabilizes.



Y	Z	AA	AB	AC	AD	AE	AF	AG	AH	AI	AJ	AK	AL	AM	AN	AO	AP	AQ	
1	Ci	gs	VPD	A	E	WUE	rb	StomataR	ZDiff	Ccontrol	Hcontrol	Tsensor	Tcontrol	Lcontrol	Accessory	Status	empty A	actual A	Ci
2	88.1632	826.533	0.81187	1.94581	5.07638	0.38331	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.2143	2.1601	87.7053
3	88.1632	826.533	0.81187	1.94581	5.07638	0.38331	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.16965	2.11545	87.8147
4	88.3847	826.354	0.80894	1.88421	5.05704	0.37259	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.13647	2.02068	88.1173
5	88.4868	812.194	0.81666	1.84033	5.03937	0.36519	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.13038	1.97071	88.164
6	88.5127	815.089	0.81386	1.83704	5.03547	0.36482	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.09949	1.93653	88.3322
7	88.5685	803.093	0.82173	1.81536	5.0277	0.36107	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.10562	1.92098	88.37
8	88.6505	798.93	0.82426	1.80022	5.02329	0.35837	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.10896	1.90917	88.3775
9	88.6204	791.734	0.82657	1.79844	5.00274	0.35949	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.10621	1.90465	88.2912
10	88.4585	779.332	0.83544	1.81092	4.99617	0.36246	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.102	1.91291	88.1319
11	88.3076	774.346	0.83923	1.833	4.99428	0.36702	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.102	1.93499	88.0487
12	88.3008	770.971	0.83889	1.83103	4.9755	0.36801	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.07088	1.90191	88.1865
13	88.3733	761.071	0.84501	1.80767	4.96236	0.36428	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.04113	1.848	88.1011
14	88.6902	748.596	0.8528	1.73388	4.94502	0.35063	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.04234	1.7762	88.1011
15	88.6431	742.874	0.85504	1.74093	4.92868	0.35322	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.05032	1.7912	88.1011
16	88.404	728.91	0.86531	1.7747	4.91521	0.36106	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	-0.04365	1.8183	88.1011
17	88.5593	725.082	0.86636	1.75814	4.90112	0.35872	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	0.00523	1.7529	88.1011
18	88.4598	723.613	0.86741	1.81596	4.89924	0.37066	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	0.09421	1.7217	88.1011
19	87.7564	703.972	0.8829	1.99927	4.88124	0.40958	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	0.31881	1.6804	88.1011
20	87.3476	707.609	0.87863	2.22791	4.87693	0.45683	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	0.65164	1.5762	88.1011
21	86.5706	703.163	0.88256	2.5869	4.87482	0.53067	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	1.13546	1.4514	88.1011
22	85.7043	706.819	0.87705	3.03097	4.86384	0.62316	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	1.61891	1.4120	88.1011
23	84.5587	696.616	0.88607	3.54046	4.85849	0.72872	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	2.08032	1.4601	88.1011
24	83.3388	680.399	0.89892	4.0564	4.839	0.83827	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	2.53269	1.5237	88.1011
25	82.2797	674.573	0.90245	4.78035	4.82515	0.99072	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	3.08009	1.7002	88.1011
26	81.8839	666.801	0.9088	5.2308	4.81495	1.08637	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	3.52729	1.70351	93.0508
27	81.4851	664.13	0.90833	5.66711	4.79697	1.18139	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	3.95636	1.71074	94.0046
28	80.9036	647.482	0.92384	6.07185	4.78188	1.26976	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	4.55668	1.51517	95.5812
29	80.8395	646.286	0.91907	6.46346	4.74978	1.36079	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	4.90713	1.55633	96.7814
30	81.0793	636.965	0.92805	6.74176	4.74105	1.422	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	5.18082	1.56094	98.0977
31	81.342	628.291	0.93537	7.03579	4.7264	1.48861	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	5.43151	1.60427	99.2982
32	81.6165	625.237	0.93505	7.37476	4.70628	1.56701	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	5.68298	1.69179	100.544
33	81.952	616.238	0.94447	7.6693	4.69905	1.6321	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	5.86286	1.80644	101.789
34	82.7061	606.891	0.94769	7.86582	4.6574	1.68889	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	6.08198	1.78384	103.465
35	83.3151	597.179	0.95647	8.09492	4.63978	1.74468	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	6.30077	1.79415	105.163
36	84.4016	595.877	0.95613	8.31106	4.62978	1.79513	0.4	0.5	AZSDB	LR	F%	IR	CT	LED	2	ML	6.41071	1.90036	106.645

Plot actual A (row AP) vs actual C_i (row AQ) starting at row 20

Sample Results

A sample linear ramp experiment was recorded using a ramp from 100 to 900 $\mu\text{mol mol}^{-1}$ with a ramp step of 150 $\mu\text{mol mol}^{-1} \text{min}^{-1}$ and cuvette flow of 250 ml/min. The CIRAS-4 was warmed up and a Stored Diff Balance Calibration was performed. Once the Stored Diff Bal was complete, a ramp data set was recorded with an empty chamber. Next, a second ramp data set was recorded with an *Ocimum basilicum* leaf and PAR set to 1000.

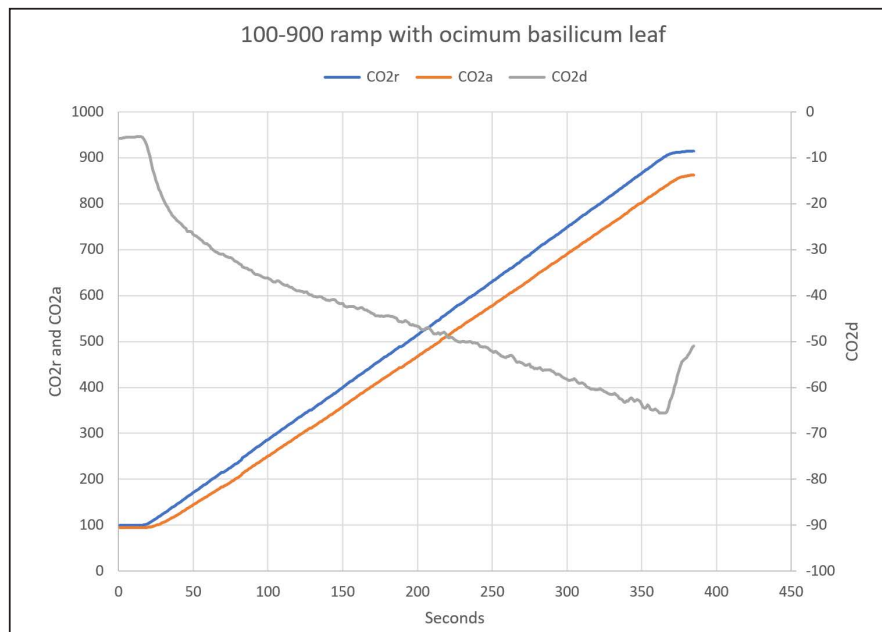
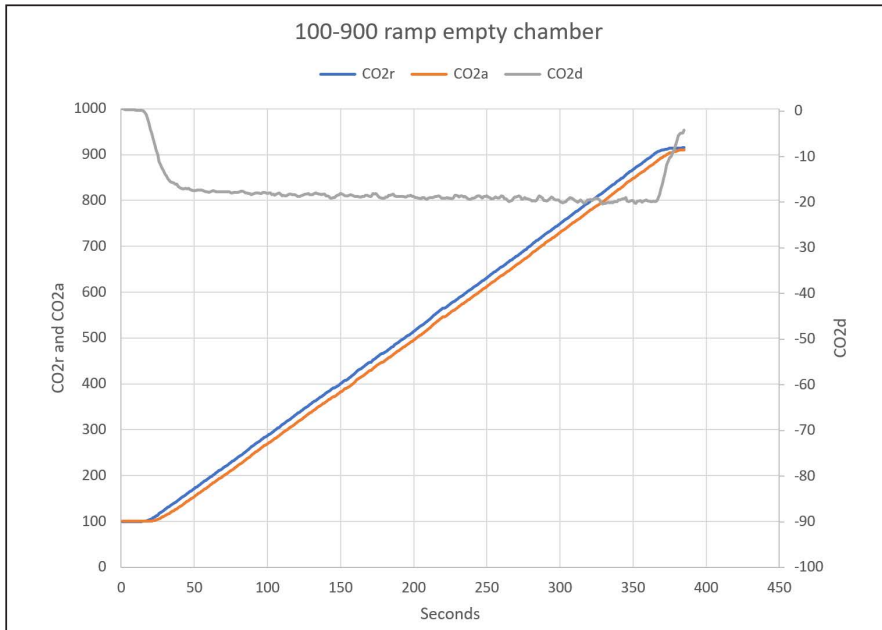
In the "no-leaf" case, the CO₂ differential is a relatively constant -20 $\mu\text{mol mol}^{-1}$ for most of the ramp, after starting out at 0 $\mu\text{mol mol}^{-1}$. The -20 $\mu\text{mol mol}^{-1}$ CO₂d represents the response time of the system including cuvette mixing and gas transport back to the CIRAS-4 console, equivalent to 13 sec with these particular settings. Faster response time can be obtained with higher cuvette flow rate, however the corresponding CO₂ differential will be lower. The CIRAS-4's

ability to perform a Stored Diff Balance over the full range of the ramp prior to running the response script eliminates the need to correct the reference and analysis for accumulated channel difference.

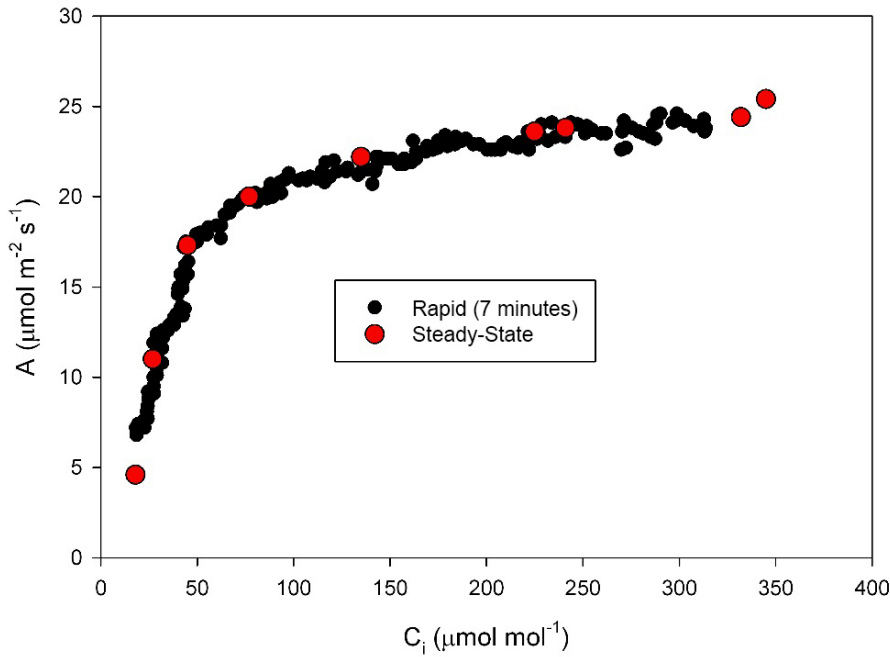
With an active leaf in the PLC4, the CO₂d begins at -6 $\mu\text{mol mol}^{-1}$ (instead of 0 as in the 'no-leaf' case) because the leaf is actively assimilating. As the CO₂r increases from 100 to 900, the CO₂d increases from -6 $\mu\text{mol mol}^{-1}$ to -65 $\mu\text{mol mol}^{-1}$ at the end of the ramp.

A vs C_i Comparisons

The Rapid A-C_i curve technique and traditional point-by-point steady-state A-C_i technique were compared on identical leaves a few minutes apart. Data on both C₃ (soybean) and a C₄ (giant foxtail) were made and show very good agreement between the two methods.

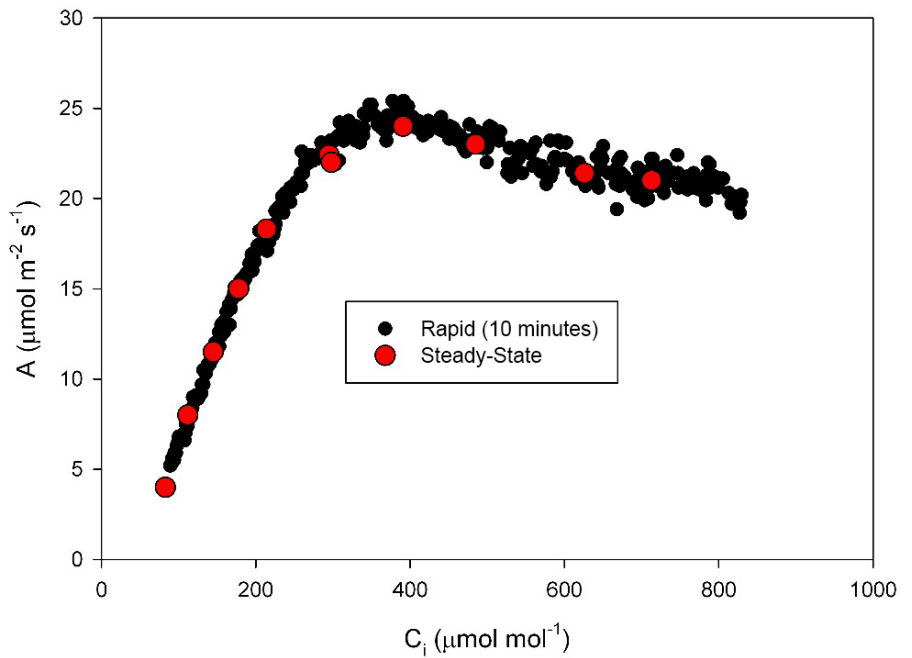


Giant Foxtail at 25°C



Comparison of High-Speed A/C_i Ramping (black points) to traditional point-by-point Steady-State (red points) for a typical C₄ Giant Foxtail leaf with PAR of 1500 μmol m⁻² s⁻¹ and Cuvette Flow of 300 ml/min. Reference CO₂ was ramped from 50 to 500 in 5 minutes (with one initial 2-minute acclimation). Each Steady-State point had a 2-minute acclimation time for total data recording time of 18 minutes.

Soybean at 25°C



Comparison of High-Speed A/C_i Ramping (black points) to traditional point-by-point Steady-State (red points) for a typical C₃ Soybean leaf with PAR of 1500 μmol m⁻² s⁻¹ and Cuvette Flow of 300 ml/min. Reference CO₂ was ramped from 100 to 1000 in 8 minutes (with one initial 2-minute acclimation). Each Steady-State point had a 2-minute acclimation time for total data recording time of 22 minutes.



Tips

1. The Stored Diff Balance Calibration should be performed after a 30-minute initial warmup prior to the beginning of a day of testing. The Stored Diff Balance will remain stable throughout a full 8-12 hour day of testing and will not be required again until the following day.
2. The *No Leaf (empty chamber)* ramp [see step 5] is stable for four hours or more and does not need to be redone unless experimental parameters are changed.
3. We have not encountered a situation where humidity values or g_s values change rapidly enough during the ramp to cause substantial errors in g_s and calculated C_i . A ramp speed of 100 to 230 $\mu\text{mol mol}^{-1}$ per minute allows curves up to saturating A to be completed in about five minutes, and g_s changes with C_i are typically minimal during that time.
4. Ramp speeds up to 233 ppm per minute are acceptable. Changing the speed of ramping in the range of 100 to 230 $\mu\text{mol mol}^{-1}$ per minute did not have a significant effect on the results in tests with *Abutilon theophrasti* (C_3). For typical C_3 leaves, a rate of 150 $\mu\text{mol mol}^{-1}$ per minute, starting at 100 $\mu\text{mol mol}^{-1}$ gives a complete A vs. C_i curve in about five minutes, due to the fact that it has been our experience that saturating CO_2 is often 800-900 $\mu\text{mol mol}^{-1}$.
5. When executing multiple successive ramps, periodically perform a manual "zero" to keep the Auto-Zero function that takes place every 30 minutes from interrupting your CO_2 ramp.
6. When performing ramps at a range of temperatures, we recommend setting the chamber temperature high enough to prevent condensation at 100% RH (i.e. above the current ambient temperature). We have also found that when performing ramps at a range of temperatures (common in field situations), the initial Stored Diff Bal should be performed with the cuvette temperature set high enough to prevent condensation at 100% RH (i.e. above the current ambient temperature). This will help to avoid warning messages associated with excessive humidity particularly outdoors where the humidity is generally much higher than indoors.
7. For most purposes, ramping up in CO_2 is a more efficient way to collect information on V_c , J and TPU from A vs. C_i curves compared to ramping down. When ramping CO_2 upwards, it is best to equilibrate the leaf to approximately 100 $\mu\text{mol mol}^{-1}$ CO_2 , using 100 $\mu\text{mol mol}^{-1}$ as the starting CO_2 level of the ramp. This will keep Rubisco activated and have stomata open. This is more efficient because the upward ramp can be terminated once the apparent photosynthesis rate is no longer increasing as CO_2 goes up, as opposed to letting the ramp go to completion—saving considerable time on each ramp. Ramping down does provide more information at lower C_i values than does ramping upward, if that information is of particular interest. However, the drawback to ramping downward is not knowing what high level of CO_2 is appropriate to start. If the starting CO_2 is too high, time is wasted. If it's too low, valuable information about the CO_2 level which saturates photosynthesis is lost. Both can be learned by trial and error, but that also takes time.




If you would like to learn more about this application or speak with one of our experienced technical staff, please feel free to get in direct contact with us via any of the contact information listed below:

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